

**AQUATIC ANIMAL DISEASE REPORT - 2021**

Country/territory: **PHILIPPINES**

Item	Disease status/occurrence code a/b/												Level of diagnosis	Epidemiological comment numbers
	Month													
FINFISH DISEASES	January	February	March	April	May	June	July	August	September	October	November	December		
<b>OIE-listed diseases</b>														
1. Infection with epizootic haematopoietic necrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
2. Infection with infectious haematopoietic necrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
3. Infection with spring viremia of carp virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
4. Infection with viral haemorrhagic septicaemia virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
5. Infection with <i>Aphanomyces invadans</i> (EUS)	-	-	-	-	-	-	-	-	-	-	-	-	I	1
6. Infection with red sea bream iridovirus	-	-	-	-	-	-	-	-	-	-	-	-	I, III	2
7. Infection with koi herpesvirus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
<b>Non OIE-listed diseases</b>														
8. Grouper iridoviral disease	-	-	-	-	-	-	-	-	-	-	-	-		
9. Viral encephalopathy and retinopathy	-	-	-	-	-	-	-	-	-	-	-	-	I, III	3
10. Enteric septicaemia of catfish	***	***	***	***	***	***	***	***	***	***	***	***		
11. Carp Edema Virus Disease	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
12. Tilapia lake virus (TiLV)	+	+	-	+	+	-	-	+	+	+	+	-	I, III	4
<b>MOLLUSC DISEASES</b>														
<b>OIE-listed diseases</b>														
1. Infection with <i>Bonamia exitiosa</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
2. Infection with <i>Perkinsus olseni</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
3. Infection with abalone herpesvirus	***	***	***	***	***	***	***	***	***	***	***	***		
4. Infection with <i>Xenohalotis californiensis</i>	***	***	***	***	***	***	***	***	***	***	***	***		
5. Infection with <i>Bonamia ostreae</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
<b>Non OIE-listed diseases</b>														
6. Infection with <i>Marteiloides chungmuensis</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
7. Acute viral necrosis (in scallops)	***	***	***	***	***	***	***	***	***	***	***	***		
<b>CRUSTACEAN DISEASES</b>														
<b>OIE-listed diseases</b>														
1. Infection with Taura syndrome virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	I, III	5
2. Infection with white spot syndrome virus	+	+	+	+	+	+	+	+	+	+	+	+	I, III	6
3. Infection with yellow head virus genotype 1	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	I, III	7
4. Infection with infectious hypodermal and haematopoietic necrosis virus	+	+	+	+	+	+	-	+	+	+	+	-	I, III	8
5. Infection with infectious myonecrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	I, III	9
6. Infection with <i>Macrobrachium rosenbergii</i> nodavirus (White Tail disease)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	I, III	
7. Infection with <i>Hepatobacter penaei</i> (Necrotising hepatopancreatitis)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	I, III	10
8. Acute hepatopancreatic necrosis disease (AHPND)	+	+	+	+	-	+	+	+	+	+	+	+	I, III	11
9. Infection with <i>Aphanomyces astaci</i> (Crayfish plague)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
<b>Non OIE-listed diseases</b>														
10. Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i> (HPM-EHP)	+	+	+	+	-	-	+	+	+	+	+	+	I, III	12
11. Viral covert mortality disease (VCMD) of shrimps	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
12. <i>Spiroplasma eriocheiris</i> infection	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
13. Decapod iridescent virus 1 (DIV-1)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000		
<b>AMPHIBIAN DISEASES</b>														
<b>OIE-listed diseases</b>														
1. Infection with <i>Ranavirus</i> species	***	***	***	***	***	***	***	***	***	***	***	***		
2. Infection with <i>Batrachochytrium dendrobatidis</i>	***	***	***	***	***	***	***	***	***	***	***	***		
3. Infection with <i>Batrachochytrium salamandrivorans</i>	***	***	***	***	***	***	***	***	***	***	***	***		

Prepared by:

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 Position: Veterinarian IV  
 Date: December 15, 2021

ANY OTHER DISEASES OF IMPORTANCE												
1												
2												

**DISEASES PRESUMED EXOTIC TO THE REGION<sup>b</sup>**  
**LISTED BY THE OIE**  
**Finfish:** Infection with HPR-deleted or HPR0 salmon anaemia virus; Infection with salmon pancreas disease virus; Infection with *Gyrodactylus salaris*.  
**Molluscs:** Infection with *Marteilia refringens*; *Perkinsus marinus*.  
**NOT LISTED BY THE OIE**  
**Finfish:** Channel catfish virus disease

a/ Please use the following occurrence code:

<u>Occurrence code and symbol</u>	<u>Definition</u>	<u>Occurrence code and symbol</u>	<u>Definition</u>
Disease present +	The disease is present with clinical signs in the whole country (in domestic species or wildlife)	Disease absent -	The disease was absent in the country during the reporting period (in domestic species or wildlife).
Disease limited to one or more zones +()	The disease is present with clinical signs, and limited to one or more zones/compartments (in domestic species or wildlife)	Never reported 0000	The disease has "never been reported" (historically absent) for the whole country in domestic species and wildlife.
Infection/infestation +?	Confirmed infestation or infection using diagnostic tests, but no clinical signs observed (in domestic species or wildlife)	No information ***	No information is available regarding the presence or the absence of this disease during the reporting period (in domestic species or wildlife).
Infection/infestation limited to one or more zones +?()	Confirmed infestation or infection using diagnostic tests, but no clinical signs observed and limited to one or more zones/compartments (in domestic species or wildlife)		
Disease suspected ?	The presence of the disease was suspected but not confirmed (in domestic species or wildlife)		
Disease suspected but not confirmed and limited to one or more zones ?()	The presence of the disease was suspected but not confirmed and limited to one or more zones/compartments (in domestic species or wildlife)		

b/ If there is any changes on historical data, please highlight in RED

**1. Epidemiological comments:**  
 (Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	<p><b>Infection with <i>Aphanomyces invadans</i> (EUS)</b>  <u>First Quarter:</u>            EUS was not detected by gross morphological examination in <i>Anguilla</i> spp. (elver), <i>A.bicolor</i> (elver) and <i>A. bicolor pacifica</i>(juvenile) from Nueva Ecija, Batangas and Tarlac. Examinations were conducted by BFAR Central and Regional Laboratories.</p> <p><u>Second Quarter:</u>            EUS was not detected by gross morphological examination in <i>Anguilla</i> spp. (grow-out) from Batangas and Cabadbaran. Examination was conducted by BFAR Central Fish Health Laboratory.</p> <p><u>Third Quarter:</u>            EUS was not detected by gross morphological examination in Eel (grow-out) and <i>Anguilla bicolor</i> (grow-out) from Batangas, Cavite and Pampanga. Examination was conducted by BFAR Central Fish Health Laboratory.</p> <p><u>Fourth Quarter:</u>            EUS was not detected by gross morphological examination in Eel (grow-out) and <i>Anguilla bicolor</i> (grow-out) from Batangas. Examination was conducted by BFAR Central Fish Health Laboratory.</p>

2	<p><b><u>Red Seabream Iridoviral Disease (RSID)</u></b>  <b><u>First Quarter:</u></b>  Tilapia (fry) and Pompano (grow-out) analyzed using PCR test showed negative results for Red Seabream Iridoviral Disease. Samples were collected from Iloilo and Nueva Ecija. Examinations were conducted by BFAR Central, Regional and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><b><u>Second Quarter:</u></b>  <i>Siganus guttatus</i>, Milkfish (fingerling and grow-out) and Tilapia (fingerling and grow-out) analyzed using PCR test showed negative results for Red Seabream Iridoviral disease. Samples were collected from Iloilo, Davao del Sur, Davao City, Nueva Ecija and Batangas. Examination was conducted by SEAFDEC and BFAR Central Fish Health Laboratory.</p> <p><b><u>Third Quarter:</u></b>  <i>Decapterus macrossoma</i>, Pompano (fingerlings), Molobicus Saline Tilapia (fingerlings), Milkfish (grow-out), and Tilapia (fingerlings and grow-out) were analyzed using Polymerase Chain Reaction (PCR) test showed negative results for Red Seabream Iridoviral disease. Samples were collected from Iloilo, Dagupan, Agusan del Norte, Saranggani Province, Rizal, Nueva Ecija, and Davao Occidental. Examination was conducted by BFAR Central and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><b><u>Fourth Quarter:</u></b></p>
3	<p><b><u>Viral Encephalopathy and Retinopathy (VER)</u></b>  <b><u>First Quarter:</u></b>  Pompano (fingerlings, grow-out), tilapia (fry) analyzed using PCR showed negative results for Viral Encephalopathy and Retinopathy. Samples were collected from Iloilo and Nueva Ecija. Examinations were conducted by BFAR Central and (SEAFDEC) Fish Health Laborator</p> <p><b><u>Second Quarter:</u></b>  Milkfish (fingerling and grow-out), <i>Siganus guttatus</i>, and Tilapia (fingerling and grow-out) analyzed using PCR tests showed negative results for Viral Encephalopathy and Retinopathy. Samples were collected from Iloilo, Davao City, Nueva Ecija and Batangas. Examinations were conducted by SEAFDEC and BFAR Central Fish Health Laboratories.</p> <p><b><u>Third Quarter:</u></b>  <i>Decapterus macrossoma</i>, Pompano (fingerlings), Grouper (grow-out), Milkfish (grow-out), Molobic Saline Tilapia (fingerlings), and Tilapia (fingerlings, juvenile and grow-out) were analyzed using Polymerase Chain Reaction (PCR) test showed negative results for Viral Encephalopathy and Retinopathy. Samples were collected from Iloilo, Dagupan, Binangonan, Camarines Sur, Agusan del Norte, Marinduque, Albay, Sarangani Province, Nueva Ecija, and Davao Occidental. Examination was conducted by BFAR Central and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><b><u>Fourth Quarter:</u></b>  Milkfish (broodstock and grow out), Seabass (fingerling), 3-stripe damsel (grow out), Blue Anchas (grow out), green chromis (grow out), purple queen (grow out), Yellow anchas (grow out), yellow damsel (grow out), <i>O. niloticus</i> (fingerlings and grow out), and shortfin scad were subjected to Polymerase Chain Reaction (PCR) analysis and showed negative results for VER. Samples were collected from Eastern Samar, Zambales, Cebu, Nueva Ecija, Occidental Mindoro, Agusan del Norte, and Iloilo. Examination was conducted by BFAR Central and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p>
4	<p><b><u>Tilapia Lake Virus (TiLV)</u></b>  <b><u>First Quarter:</u></b>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected 3 farms  <b>Species affected:</b> Tilapia (fingerlings, fry, juvenile)  <b>Pathogen:</b> Tilapia Lake Virus  <b>Size of infected areas or names of infected areas:</b> La Union, Nueva Ecija and Agusan del Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories):</b> Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratory</p> <p><b><u>Second Quarter:</u></b>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 2 farms  <b>Species affected:</b> Tilapia (fingerlings, grow-out and broodstock)  <b>Pathogen:</b> Tilapia Lake Virus  <b>Size of infected areas or names of infected areas:</b> Bohol and Laguna  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central and BFAR Regional Fish Health Laboratory</p> <p><b><u>Third Quarter:</u></b>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 8 farms  <b>Species affected:</b> Tilapia (juvenile, grow-out, breeder, and adult) and <i>Oreochromis</i> spp. (grow-out)  <b>Pathogen:</b> Tilapia Lake Virus  <b>Size of infected areas or names of infected areas:</b> Bukidnon, Bohol, Lanao del Norte, Davao Occidental, and Agusan del Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central and BFAR Regional Fish Health Laboratories</p> <p><b><u>Fourth Quarter:</u></b>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 2 farms</p>
5	<p><b><u>Taura Syndrome (TS)</u></b>  <b><u>First Quarter:</u></b>  <i>P.vannamei</i> (broodstock, adult, grow-out, juvenile, post-larvae, fry and nauplii) analyzed using PCR test showed negative for Taura Syndrome. Samples were collected from Butuan, Iloilo, Bulacan, Leyte, Oriental Mindoro, Cagayan, Pangasinan, Zambales, Quezon, Sorsogon, Camarines Norte, Bohol and Cebu. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories.</p> <p><b><u>Second Quarter:</u></b>  <i>P. monodon</i> (grow-out and post-larvae) and <i>P. vannamei</i> (fry, nursery, post-larvae, adult, grow-out and broodstock) analyzed using PCR tests showed negative for Taura Syndrome. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Zambales, Bohol, Pangasinan, Nueva Ecija, Quezon, and Iloilo. Examinations were conducted by BFAR Central and BFAR Regional Fish Health Laboratories.</p> <p><b><u>Third Quarter:</u></b>  <i>P. monodon</i> (post-larvae and grow-out), <i>P. indicus</i> and <i>P. vannamei</i> (nauplii, post larvae, grow-out, adult, and broodstock) were analyzed using Polymerase Chain Reaction (PCR) test showed negative results for Taura Syndrome. Samples were collected from Bulacan, Oriental Mindoro, Occidental Mindoro, Surigao City, Zambales, Cebu, Agusan del Norte, Butuan, Davao del Sur, Leyte, Pangasinan, Quezon, Batangas, and Bohol. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories.</p>

6	<p><b>White Spot Disease (WSD)</b>  <u>First Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected 3 farms  <b>Species affected:</b> <i>P.monodon</i> (broodstock and post-larvae) and <i>P.vannamei</i> (grow-out and juvenile)  <b>Pathogen:</b> White Spot Syndrome Virus  <b>Size infected areas or names of infected areas:</b> Negros Occidental, Leyte, Cagayan, Mindoro, Sorsogon, Camarines Norte, Davao del Sur, Agusan del Norte, Misamis Oriental, Surigao del Sur  <b>Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories):</b> Polymerase Chain Reaction (PCR) Test / BFAR Central, Regional and SEAFDEC Fish Health Laboratories.</p> <p><u>Second Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 17 farms  <b>Species affected:</b> <i>P. Vannamei</i> (post-larvae, grow-out and spawner), <i>P. monodon</i> (post-larvae, grow-out and spawner), <i>S. olivacea</i> (grow-out) and <i>P. indicus</i> (grow-out)  <b>Pathogen:</b> White Spot Syndrome Virus  <b>Size of infected areas or names of infected areas</b> : Iloilo, Negros Occidental, Oriental Mindoro, Cagayan, Agusan del Norte, Sorsogon, Camarines Norte, Cebu, Leyte and Leyte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction (PCR), Conventional, and IQ Plus tests/ BFAR Central, BFAR Regional and SEAFDEC Fish Health Laboratories.</p> <p><u>Third Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 10 farms  <b>Species affected:</b> <i>P. vannamei</i> (post-larvae and grow-out), <i>P. monodon</i> (post-larvae and grow-out), and <i>S. serrata</i> (broodstock)  <b>Pathogen:</b> White Spot Syndrome Virus  <b>Size of infected areas or names of infected areas:</b> Iloilo, Capiz, Camarines Sur, Leyte, and Agusan del Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction (PCR) test / BFAR Regional and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><u>Fourth Quarter:</u></p>
7	<p><b>Infection with Yellow Head Virus Genotype 1 (YHV)</b>  <u>First Quarter:</u>  <i>P.monodon</i> ( fry and grow-out) and <i>P.vannamei</i> (broodstock and grow-out) analyzed using PCR test showed negative results for Yellow Head Virus Genotype 1. Samples were collected from Butuan, Iloilo, Bulacan, Leyte and Mindoro. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories.</p> <p><u>Second Quarter:</u>  <i>P. monodon</i> (post-larvae, grow-out and broodstock) and <i>P. vannamei</i> (post-larvae, grow-out, adult and broodstock) analyzed using Conventional test showed negative result for Yellow Head Virus. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Zambales, Pangasinan, Bohol, Iloilo and Misamis Occidental. Examination was conducted by BFAR Central Fish and BFAR Regional Health Laboratories.</p> <p><u>Third Quarter:</u>  <i>P. monodon</i> (grow-out), <i>P. vannamei</i> (post-larvae and broodstock), and <i>P. indicus</i> were analyzed using Polymerase Chain Reaction (PCR) test showed negative result for Yellow Head Virus. Samples were collected from Bulacan, Oriental Mindoro, Occidental Mindoro, Leyte, Surigao City, Zambales, Cebu, Agusan del Norte, Butuan, and Davao del Sur. Examination was conducted by BFAR Central Fish and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p>
8	<p><b>Infection with Infectious Hypodermal and Haematopoietic Necrosis Virus (IHNV)</b>  <u>First Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected 6 farms  <b>Species affected:</b> <i>P.monodon</i> (post larvae) and <i>P.vannamei</i> (post-larvae, adult)  <b>Pathogen:</b> Infectious Hypodermal and Haematopoietic Necrosis Virus  <b>Size infected areas or names of infected areas:</b> Pampanga, Zambales, Quezon, Camarines Norte, Bohol and Cebu  <b>Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories):</b> Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratories.</p> <p><u>Second Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 9 farms  <b>Species affected:</b> <i>P. monodon</i> (post-larvae and grow-out) and <i>P. vannamei</i> (fry and adult)  <b>Pathogen:</b> Infectious Hypodermal and Haematopoietic Virus  <b>Size of infected areas or names of infected areas</b> : Bulacan, Zambales, Sorsogon, and Camarines Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction (PCR) / BFAR Central and BFAR Regional Fish Health Laboratories.</p> <p><u>Third Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 14 farms  <b>Species affected:</b> <i>P. monodon</i> (post-larvae), <i>P. indicus</i> and <i>P. vannamei</i>  <b>Pathogen:</b> Infectious Hypodermal and Haematopoietic Virus  <b>Size of infected areas or names of infected areas:</b> Oriental Mindoro, Surigao City, Occidental Mindoro, and Camarines Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction (PCR) test / BFAR Central and Regional Fish Health Laboratories.</p> <p><u>Fourth Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 2 farms  <b>Species affected:</b> <i>P. vannamei</i> (post-larvae and grow out)  <b>Pathogen:</b> Infectious Hypodermal and Haematopoietic Virus  <b>Size of infected areas or names of infected areas:</b> Oriental Mindoro, Surigao City, Occidental Mindoro, and Camarines Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction (PCR) test / BFAR Central and Regional Fish Health Laboratories.</p>

9	<p><b>Infectious Myonecrosis (IMN)</b></p> <p><u>First Quarter:</u>  <i>P.monodon</i> ( fry, grow-out) and <i>P.vannamei</i> (broodstock, grow-out, adult) analyzed using PCR test showed negative for Infectious Myonecrosis. Samples were collected from Butuan, Iloilo, Bulacan, Leyte, Mindoro, Cagayan, Bohol and Cebu. Examinations were conducted by BFAR Regional Fish Health Laboratory.</p> <p><u>Second Quarter:</u>  <i>P. monodon</i> (post-larvae, grow-out and adult) and <i>P. vannamei</i> (nursery, post-larvae, grow-out, broodstock and breeders) analyzed using PCR tests showed negative for Infectious Myonecrosis. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Pangasinan, Zambales, Quezon, Iloilo and Bohol. Examinations were conducted by BFAR Central and BFAR Regional Fish Health Laboratories.</p> <p><u>Third Quarter:</u>  <i>P. monodon</i> (post-larvae and grow-out), <i>P. vannamei</i> (post-larvae, grow-out, and broodstock), and <i>P. indicus</i> were analyzed using Polymerase Chain Reaction (PCR) test showed negative for Infectious Myonecrosis. Samples were collected from Iloilo, Bulacan, Oriental Mindoro, Occidental Mindoro, Quezon, Surigao City, Zambales, Agusan del Norte, Butuan City, Bohol, Batangas, Davao del Sur, Leyte, and Cebu. Examinations were conducted by BFAR Central, Regional, and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><u>Fourth Quarter:</u>  <i>P. monodon</i> (post-larvae and grow-out), <i>P. vannamei</i> (post-larvae, grow-out, adult, and broodstock), <i>Macrobrachium rosenbergii</i>(adult), and <i>Thenus orientalis</i>analyzed using Polymerase Chain Reaction (PCR) test showed negative for Infectious Myonecrosis. Samples were collected from Bulacan, Agusan del Norte, Oriental Mindoro, Cebu, Bohol, Iloilo, Pangasinan, Cagayan, Batangas, and Quezon. Examinations were conducted by BFAR Central, Regional, and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p>
10	<p><b>Necrotising Hepatopancreatitis (NHP)</b></p> <p><u>First Quarter:</u>  <i>P.monodon</i> (post-larvae, grow-out) and <i>P.vannamei</i> (broodstock, grow-out, post-larvae) analyzed using PCR test showed negative for Necrotising Hepatopancreatitis. Samples were collected from Butuan, Iloilo, Bulacan, Leyte, Mindoro, Quezon, Bohol and Cebu. Examinations were conducted by BFAR Central Fish Health Laboratory.</p> <p><u>Second Quarter:</u>  <i>P. vannamei</i> (post-larvae, breeders and broodstock) and <i>P. monodon</i> (grow-out) analyzed using PCR tests showed negative for Necrotising Hepatopancreatitis. Samples were collected from Bulacan, Cagayan, Oriental Mindoro, Bohol and Cebu. Examinations were conducted by BFAR Central Fish Health Laboratory.</p> <p><u>Third Quarter:</u>  <i>P. vannamei</i> (broodstock), <i>P. monodon</i> (grow-out), and <i>P. indicus</i> were analyzed using Conventional test showed negative for Necrotising Hepatopancreatitis. Samples were collected from Bulacan, Butuan, Surigao del Sur, Oriental Mindoro, Occidental Mindoro, Surigao City, Zambales, Cebu, Agusan del Norte, Butuan, Davao del Sur, and Leyte. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories</p> <p><u>Fourth Quarter:</u>  <i>P. vannamei</i> (grow out and broodstock), and <i>P. monodon</i> (grow-out) analyzed using Conventional PCR test showed negative for Necrotising Hepatopancreatitis. Samples were collected from Bulacan, Agusan del Norte, Oriental Mindoro, Occidental Mindoro, Davao Oriental, Cebu, Negros Oriental, Pangasinan, Batangas, and Quezon. Examinations were conducted by BFAR Central laboratory.</p>

11	<p><b>Acute Hepatopancreatic Necrosis Disease (AHPND)</b>  <u>First Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected 5 farms  <b>Species affected:</b> <i>P.vannamei</i> (post-larvae, grow-out)  <b>Pathogen:</b> Acute Hepatopancreatic Necrosis Disease  <b>Size infected areas or names of infected areas:</b> Negros Occidental, Cagayan, Occidental Mindoro, Oriental Mindoro and Cebu  <b>Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories):</b> Polymerase Chain Reaction (PCR) Test / BFAR Regional and SEAFDEC Fish Health Laboratories.</p> <p><u>Second Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 2 farms  <b>Species affected:</b> <i>P. vannamei</i> (post-larvae and grow-out)  <b>Pathogen:</b> AHPND <i>Vibrio parahaemolyticus</i>  <b>Size of infected areas or names of infected areas:</b> Oriental Mindoro and Leyte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories)</b> : Polymerase Chain Reaction Test (PCR) / BFAR Regional Fish Health Laboratory.</p> <p><u>Third Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 9 farms  <b>Species affected:</b> <i>P. vannamei</i> (post-larvae and grow-out)  <b>Pathogen:</b> AHPND <i>Vibrio parahaemolyticus</i>  <b>Size of infected areas or names of infected areas:</b> Iloilo, Bulacan, Cagayan, Zambales, Oriental Mindoro, and Leyte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central, Regional, and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><u>Fourth Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 12 farms  <b>Species affected:</b> <i>P. vannamei</i> (grow-out and juvenile), <i>P. monodon</i> (juvenile and adult), and <i>P. indicus</i>  <b>Pathogen:</b> AHPND <i>Vibrio parahaemolyticus</i>  <b>Size of infected areas or names of infected areas:</b> Oriental Mindoro, Zambales, Cebu, Iloilo, Pangasinan, Cagayan, and Batangas  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central fish health laboratory and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratory.</p>
12	<p><b>Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i> (HPM-EHP)</b>  <u>First Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b>– detected 6 farms  <b>Species affected:</b> <i>P.monodon</i> (grow-out) and <i>P.vannamei</i> (fry, post-larvae, grow-out)  <b>Pathogen:</b> Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i>  <b>Size infected areas or names of infected areas:</b> Pampanga, Zambales, Quezon, Camarines Norte, Bohol and Cebu  <b>Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories):</b> Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratories.</p> <p><u>Second Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 1 farm  <b>Species affected:</b> <i>P. monodon</i> (post-larvae)  <b>Pathogen:</b> <i>Enterocytozoon hepatopenaei</i>  <b>Size of infected areas or names of infected areas:</b> Agusan del Norte  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Regional Fish Health Laboratory.</p> <p><u>Third Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 12 farms  <b>Species affected:</b> <i>P. monodon</i> (post-larvae and grow-out), <i>P. vannamei</i> (grow-out), and <i>P. indicus</i> (grow-out)  <b>Pathogen:</b> <i>Enterocytozoon hepatopenaei</i>  <b>Size of infected areas or names of infected areas:</b> Iloilo, Zambales, Batangas, Occidental Mindoro, Oriental Mindoro, and Misamis Occidental  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central, Regional, and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p> <p><u>Fourth Quarter:</u>  <b>Origin of the disease or pathogen (history of the disease)</b> – detected in 12 farms  <b>Species affected:</b> <i>P. monodon</i> (post-larvae and grow-out), <i>P. vannamei</i> (post-larvae, grow-out, and juvenile)  <b>Pathogen:</b> <i>Enterocytozoon hepatopenaei</i>  <b>Size of infected areas or names of infected areas:</b> Davao Oriental, Zambales, Cebu, Iloilo, Batangas, and Albay  <b>Samples sent to national or international laboratories for confirmation (indicate the name of laboratories):</b> Polymerase Chain Reaction Test (PCR) / BFAR Central, Regional, and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories.</p>

2. New aquatic animal health regulations introduced within past six months (with effective date):

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