

**Sub: OIE Manual of Standard for diagnostic Test and Vaccines - comments**

| S. No | Chapter   | Comments   |
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| 1.    | 3.4.14<br>Lumpy skin disease  | <p><b>Line 305-306:</b> An appropriate cell suspension (such as MDBK) is prepared from pregrown monolayers may be corrected as: An appropriate cell suspension (such as MDBK or LT) is prepared from pregrown monolayers, as some of the LSDV isolates do not grow well in MDBK cells.</p> <p><b>Line 681-683:</b> Reference Pestova et al. (2019) need to be corrected as Pestova et al. (2018).</p>  |
| 2.    | 3.1.7<br>Epizootic hemorrhagic disease (infection with epizootic hemorrhagic disease virus) | <p>Can be deleted the lines (Cytopathic effect (CPE), which occurs only in mammalian cell lines, usually appears between 2 and 7 days post-inoculation, however a blind passage may be required) as it is repeated in <b>lines 149-153</b> or modified suitably.</p> <p><b>Line no 53:</b> may be changed as White-tailed deer (<i>Odocoileus virginianus</i>) is the species most severely affected</p> <p><b>Line no 65</b> may be changed as erythema of the udder and difficulty in swallowing</p>   |
| 3.    | 3.3.4<br>Avian influenza (Infection with high pathogenicity avian influenza virus)          | <p><b>Line 7, 8 –</b> As per the recent ICTV classification of viruses, the influenza virus <i>Genera</i> has been modified as “<i>Alphainfluenzavirus</i>” for the previous designation of “<i>Influenzavirus A</i>”. Similarly, the “<i>Beta</i>influenzavirus”, “<i>Gamma</i>influenzavirus” and “<i>Delta</i>influenzavirus” in place of “<i>Influenzavirus B</i>”, “<i>Influenzavirus C</i>” and the newly added “<i>Influenzavirus D</i>” respectively, which has been amended in the draft and highlighted. (<u>ICTV Master Species List 2018b.v2</u>, <a href="https://talk.ictvonline.org/files/master-species-lists/m/msl/8266">https://talk.ictvonline.org/files/master-species-lists/m/msl/8266</a>, accessed on 11 December 2019). Currently, there are four genera under influenza virus.</p> <p><b>Line 88 –</b> Same as above for “<i>Influenzavirus A</i>”.</p> <p><b>Line 445-446 –</b> Please change “Hoffman” to “Hoffmann”</p> <p><b>Line 451–</b> Change suggested as “Amplicon” in place of “Amplificate”</p> <p><b>Line 776 -</b> any one of HA or haemagglutinin may be used.</p> |
| 4     | 3.7.4 Contagious Caprine Pleuropneumonia  | <p>The Chapter mentions about the classical etiological agent of Contagious caprine Pleuropneumonia i.e. <i>Mycoplasma capricolum</i> subsp. <i>capripneumoniae</i>. This chapter should also be incorporated with some information about the agent <i>Mycoplasma mycoides</i> subsp. <i>capri</i>, which is widely prevalent throughout the world among goat population and causing Caprine Pleuropneumonia. The symptoms and lesions are quite similar as well as parallel in both the case of disease.</p> <p>As compared to classical CCPP, the disease Caprine Pleuropneumonia is not that contagious in goats. Although <i>Mycoplasma mycoides</i> subsp. <i>capri</i> has been found to be involved</p>   |

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|   |   | <p>in MAKePS syndrome in goats and sheep but while conducting Trans-Tracheal inoculation experiments in goats, the similar symptoms and lesions could be observed to that of CCPP.</p> <p>It is suggested here to include some information and discussion about the organism <i>Mycoplasma mycoides</i> subsp. <i>capri</i> along with <i>Mycoplasma capricolum</i> subsp. <i>capripneumoniae</i></p>  |
| 5 | 3.1.10<br>Japanese Encephalitis   | <b>Line 15-18</b> intracerebral inoculation of suckling mice is more sensitive assay than cell culture and should be retained here   |
| 6 | 3.5.8<br>Equine piroplasmosis   | <p><b>Line 224-226:</b> Please add a reference of Kumar et al (2013). They also showed applicability of EMA-2 based ELISA for sero-diagnosis of <i>Theileria equi</i> infection. They applied it successfully on Indian Equine Population.</p> <p><i>Kumar S, Kumar R, Gupta AK, Yadav SC, Goyal SK, Khurana SK, Singh RK. 2013. Development of EMA-2 recombinant antigenbased enzyme-linked immunosorbent assay for seroprevalence studies of Theileria equi infection in Indian equine population. Vet Parasitol, 198 (1-2):10-17.</i></p> |
| 7 | 3.1.21<br><i>Trypanosoma evansi</i><br>infection (surra in all species) | <p><b>Line 9:</b> the word “arthropod-borne disease” may be written as “arthropod-transmitted disease” because infection is transmitted mechanically and there is no developmental stage in the vector.</p> <p><b>Line 531:</b><br/>“such as protein A (for detection of IgG) or protein G (for detection of IgM)” may be written as “such as protein A or protein G”</p> <p>It is not agreeable that Protein G is used for detection of IgM. It also binds IgG.</p>   |
| 8 | 3.7.9<br>Peste petits des ruminants (PPR)                               | <b>Line 695-697</b> Other attenuated PPR vaccine strain <b>Sungri/96</b> that have been developed in India have been shown to produce long-lasting immunity too (Saravanan <i>et al.</i> , 2010a; Sen <i>et al.</i> , 2010).   |