



Activities of the WOH Reference Laboratory for ASF at the Australian Centre for Disease Preparedness (ACDP)

David Williams

4th Consultation Meeting on ASF for Southeast Asia, Hanoi, Vietnam, 26th-28th Nov 2024

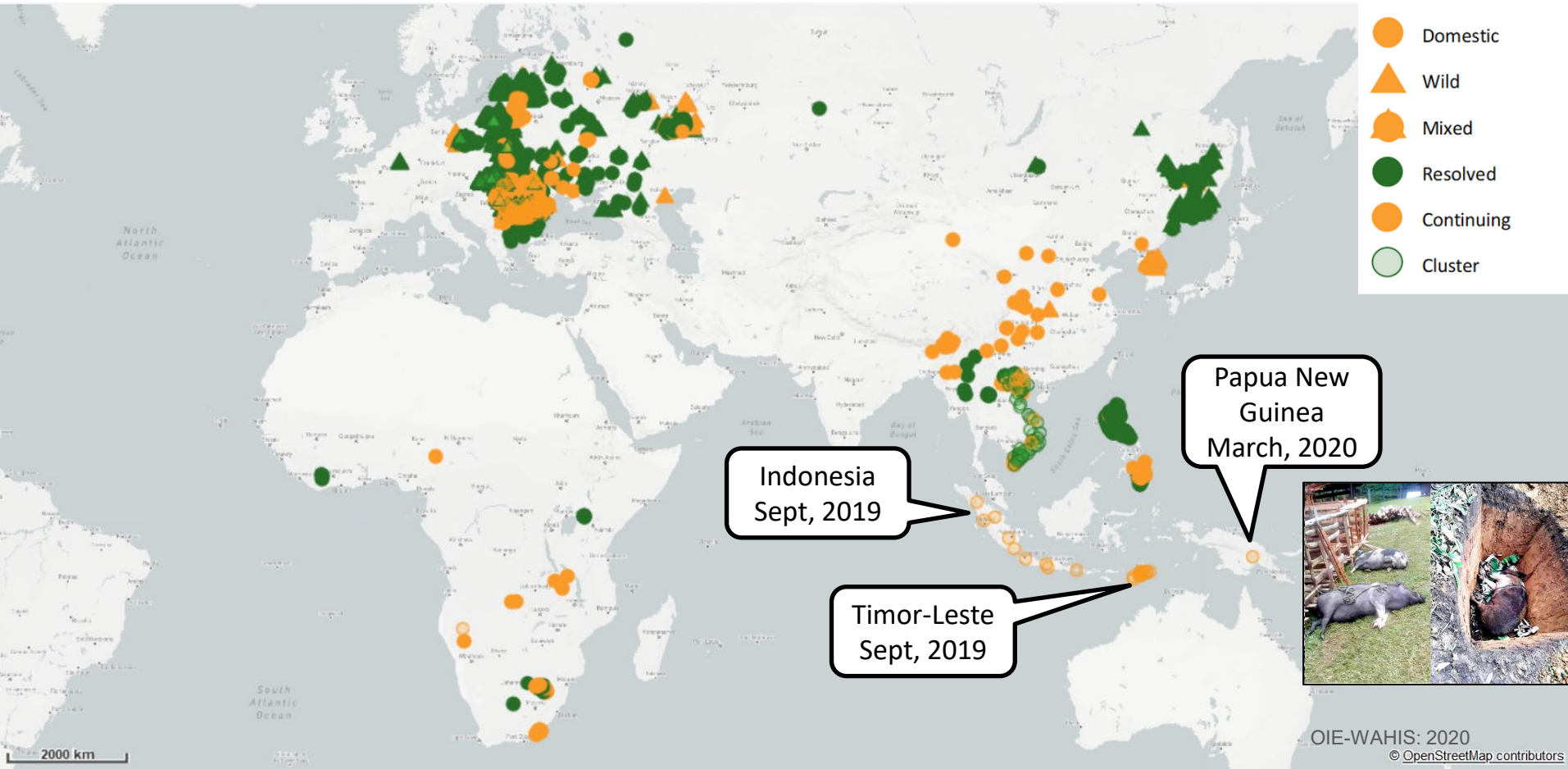
ACDP – AUSTRALIA'S NATIONAL BIOCONTAINMENT FACILITY

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ASF: on Australia's doorstep



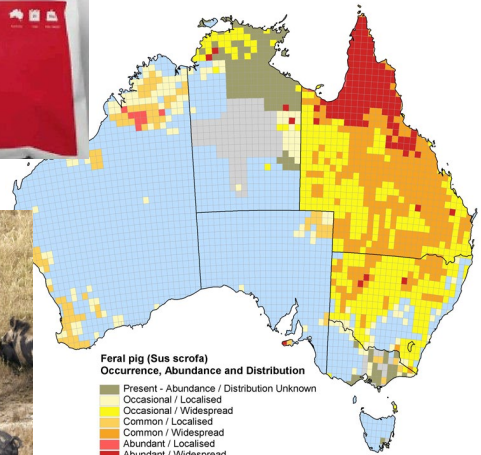
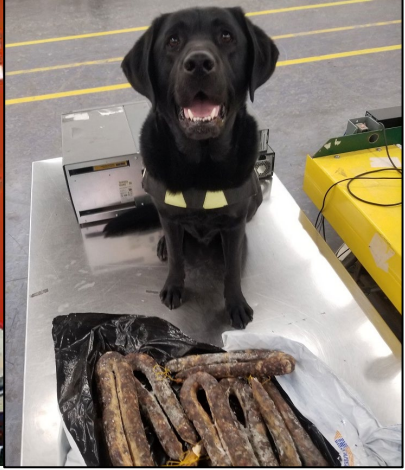


ASF threat to Australia

- Potential mechanisms of introduction
 - Infected pork and pork products
 - Contaminated porcine genetic material
 - Contaminated equipment and clothing
 - Infected pigs or carcasses of infected pigs
 - Food waste from unregulated foreign boats and shipping in Northern Australia

→ swill feeding to/contact with domestic pigs
 → eaten by free-range or feral pigs

• *Large feral pig population is potential reservoir*





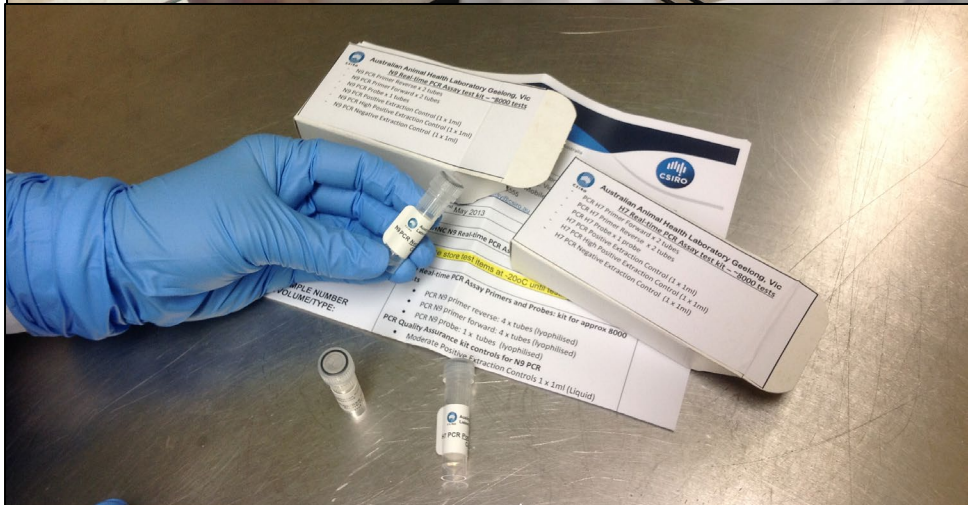
Laboratory diagnosis at ACDP

- Multidisciplinary diagnostic capability
 - Molecular (PCR and sequencing), virology, serology and pathology
- Diagnostic or confirmatory testing and virus characterisation for outbreak response and surveillance
 - Vietnam, Myanmar, Timor Leste, Papua New Guinea (PNG), Bhutan, Nepal
 - ACDP provides diagnostic and sequence analysis report
- Diagnostic capacity building for regional animal health labs
- Testing of illegally imported pork for Australian Department of Agriculture and other countries on request



Lab diagnostics

- Training and advice for:
 - Diagnostic testing (WOAH, FAO)
 - Biosafety and biosecurity
 - Sampling, packaging and shipping
- Provide diagnostic reagents (PCR kits, pos controls)
- Proficiency testing (PT): ISO/IEC 17043-accredited PT provider, coordinate a regional (Asia-Pacific) programme for swine disease, including ASF and CSF viruses





ACDP Proficiency Testing: swine diseases PCR panel

- Distributed 27 PT panels, with testing conducted in June
- Panel comprised 19 samples, including ASFV, CSFV, Influenza A, PRRSV
- 18 participating countries (no. labs):
Australia (1), Bangladesh (1), Bhutan (1), Brunei (1), Cambodia (2), Hong Kong (1), India (1), Indonesia (4), Japan (1), Korea (1), Malaysia (1), Nepal (1), Philippines (2), Singapore (1), Sri Lanka (1), Chinese Taipei (1), Thailand (3), Vietnam (8)
- Overall, the majority of laboratories returned **acceptable results for the ASF component** of the PT panel, demonstrating reliable diagnostic performance





Support for Pacific Islands Countries and Indonesia for ASF Laboratory and Field Testing

- ACDP working with Papua New Guinea National Agriculture & Quarantine Inspection Authority (NAQIA) on Laboratory Capacity Building, via 4-year project
 - Priority EADs, including ASF
 - Laboratory diagnostics (focus on PCR), biosafety and quality assurance
- Previously provided training workshops and advice for use of field-testing Indonesia and Pacific Islands Countries





Asia Pacific Placements and Laboratory Leadership (APPLLe) Project



Australian Government

Department of Foreign Affairs and Trade

- DFAT-funded project under the Partnership for a Healthy Region initiative (2023-2027)
- Can support activities throughout the Asia Pacific region with an emphasis on developing technical and leadership capability within laboratories in **Vietnam** to enable better representation in global disease networks
- *Contact: Jemma.Bergfeld@csiro.au*
- **Activities include:**
 - Laboratory placements (at ACDP or a leading laboratory within SE Asia)
 - Technical training and workshops led by ACDP staff for high priority TADs
 - Strengthening diagnostic expertise for **ASF** at RAHO6 in Ho Chi Minh City
 - Development of sequencing capability (via MinION), particularly for **ASF**
 - Co-facilitation of workshops with ACDP for other laboratories in the region
 - Development of testing protocols for investigation of new and emerging diseases at the human-animal-wildlife interface



Protocols and Guidelines for Laboratory Diagnosis

- Addresses detection of:
 - Virulent and variant forms of ASFV
 - LAVs using DIVA PCRs
 - Technical procedures and guidance
- Developed in consultation with the **WOAH ASF Reference Laboratory Network**, based on WOAH recommendations for laboratory diagnosis of ASF
- *Available online*

Addressing African Swine Fever Protocols and Guidelines for Laboratory Diagnosis

January 2024



The World Organisation for Animal Health (WOAH), the Food and Agriculture Organization of the United Nations (FAO) and other partners have been actively working in countries affected or at risk of incursion by African swine fever (ASF). This manual is an updated and expanded version of guidance first published in 2020 following the emergence of ASF in China, other Asian countries, and countries of the Pacific and Caribbean regions.



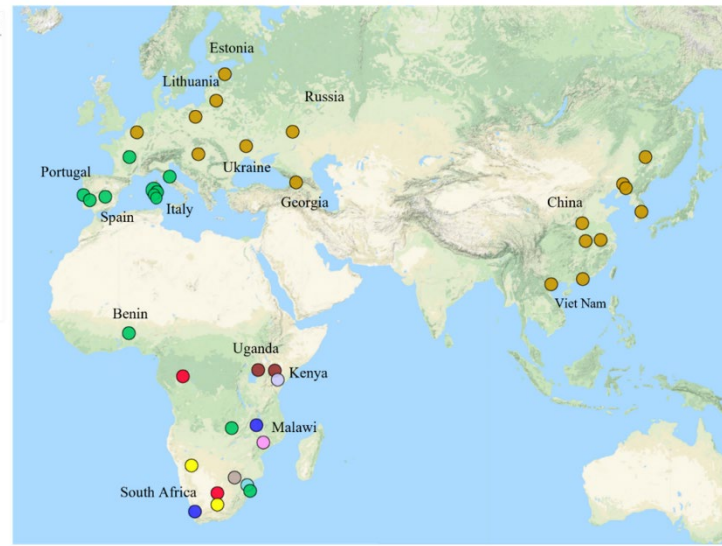
ASF molecular typing and epidemiology

- Determine genotypes of circulating viruses and genetic variation within and between populations
 - Can be used to identify recombinants
- Patterns of distribution and spread can be determined by combining surveillance data with genetic data
- Used for trace forward/trace back investigations during outbreaks, understand source and origin
- Important for vaccine matching



Molecular typing and epidemiology

- Several genes and combinations have been used
 - Partial p72 gene for genotyping, based on Bastos et al. (2003)
 - 24 genotypes
 - Only two found outside Africa
- Other targets: Central variable region of *B602L(9RL)*, *KP86R*, *E183L (p54)* and *CP204L (p30)*, *I73R-I329L* intergenic region (IGR)



IGR:

Georgia 2007/01	172351	ACAAGTATAT	GGAATATATAGGAATATATA-----	GAAATATATAGAAATAGCTAAGCT
ASFV China 2018/	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Pol14/Krus	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Bell3/Grodno	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Ukrl2/Zapo	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Lt14/1490	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Lt14/1482	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Pol14/Sz	81	ACAAGTATAT	GGAATATATAGGAATATATA	GGAATATATAGAAATAGCTAAGCT
Armenia/07	81	ACAAGTATAT	GGAATATATAGGAATATATA-----	GAAATATATAGAAATAGCTAAGCT
Tver0511/Torjo	81	ACAAGTATAT	GGAATATATAGGAATATATA-----	GAAATATATAGAAATAGCTAAGCT
Rostov09	81	ACAAGTATAT	GGAATATATAGGAATATATA-----	GAAATATATAGAAATAGCTAAGCT

Shen et al. 2022. doi.org/10.3390/v14050889



Recent Developments for ASF typing

- Full length p72 protein sequence
 - ‘proteotype’
 - 6 new p72 groups proposed:

New genotype/ proteotype	Original genotype (<i>partial p72 gene</i>)
1	I, VII, XXII
2	II, III, IV, V, XVII, XX, XXI, XXIV
8	VIII
9	IX, X
15	XV
23	XXIIIa, XXIIIb



Article

A Re-Evaluation of African Swine Fever Genotypes Based on p72 Sequences Reveals the Existence of Only Six Distinct p72 Groups

Edward Spinard ^{1,2,3}, Mark Dinhobl ^{1,2,3}, Nicolas Tesler ^{1,4}, Hillary Birtley ^{1,4}, Anthony V. Signore ^{3,5}, Aruna Ambagala ^{3,5}, Charles Maseembe ^{3,6}, Manuel V. Borca ^{1,2,3,*} and Douglas P. Gladue ^{1,2,3,*}

¹ Plum Island Animal Disease Center, Agricultural Research Service, U.S. Department of Agriculture,



Recent Developments for ASF typing

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 - **5** new p72 groups proposed:

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Virology | Announcement

African swine fever virus P72 genotyping tool

Mark Dinhobl,^{1,2,3} Edward Spinard,^{1,2,3} Hillary Birtley,^{2,3,4} Nicolas Tesler,^{2,3,4} Manuel V. Borca,^{1,2,3} Douglas P. Gladue^{1,2,3}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT Historically, genotyping of African swine fever virus was based on partial sequencing of B646L (p72). Until recently, the number of differences that defined genotypes was ambiguous. This tool allows a sequence to be uploaded and will report its closest matches along with its likely p72 genotype.

asfvgenomics.com



Recent Developments for ASF typing

- Analysis of complete protein coding sequence (*proteome*) of ASFV genome derived from whole genomes → 7 **biotypes** proposed:



Article

Reclassification of ASFV into 7 Biotypes Using Unsupervised Machine Learning

Mark Dinhob1^{1,2,3,†}, Edward Spinard^{1,2,3,†}, Nicolas Tesler^{1,4}, Hillary Birtley^{1,4}, Anthony Signore^{3,5}, Aruna Ambagala^{3,5}, Charles Masembe^{3,6}, Manuel V. Borca^{1,2,3,*} and Douglas P. Gladue^{1,2,3,*}

¹ United States Department of Agriculture, Agricultural Research Service, Foreign Animal Disease Research

Biotype	New genotype/ proteotype	Original genotype (partial p72 gene)
1	1	I, VII
1/2 R*	2	II
2	2	II
3	2	III, IV, V, XX
4	9	X
5	9	IX
6	8, 15	VIII, XV

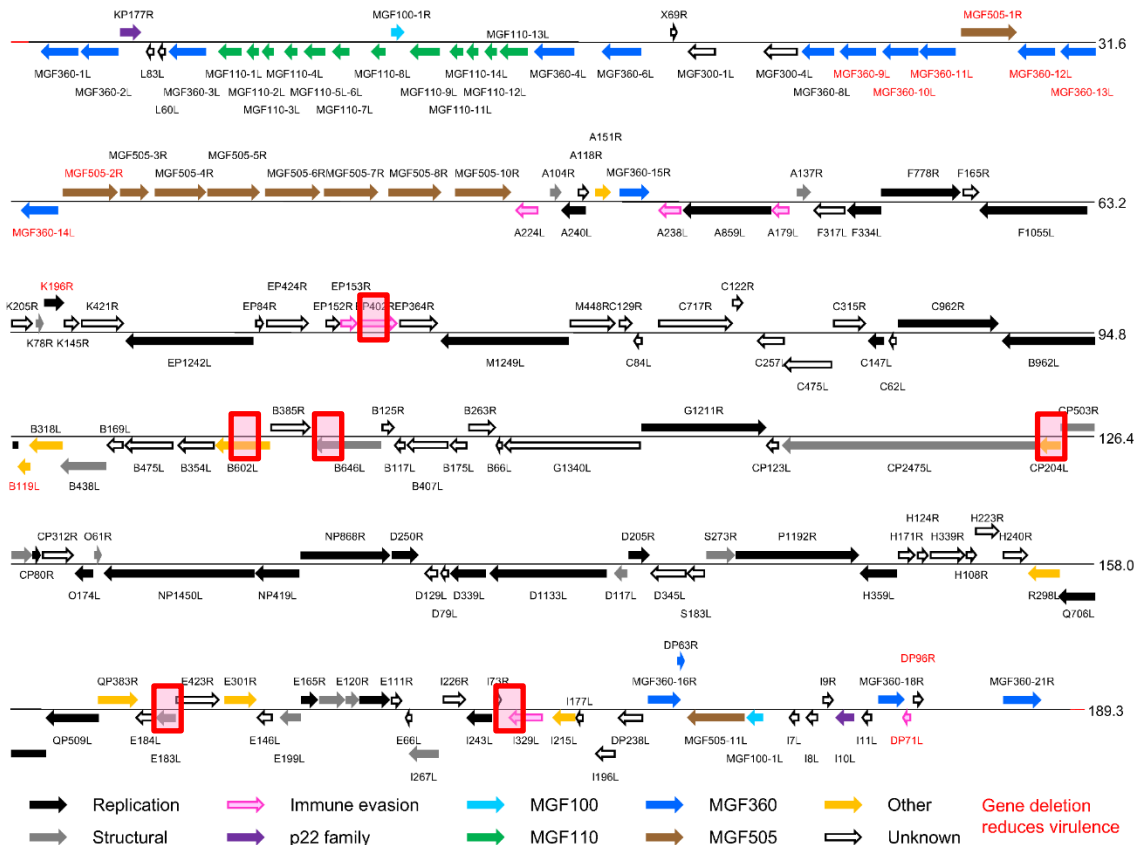
**1/2 R is comprised of recombinants genomes of biotypes 1 and 2*



A role for whole genome sequencing (WGS) for molecular epidemiology?

The ASFV genome is very large and highly stable. Individual gene targets () are of limited use for the identification of variants and the application of phylogeny for studying evolution

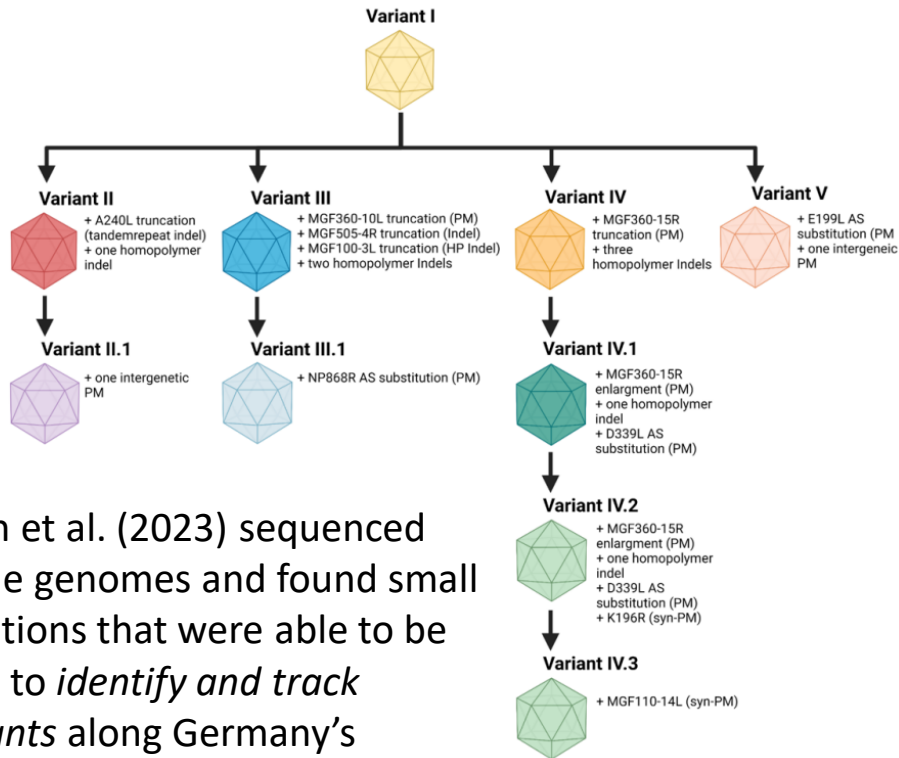
WGS are increasingly being used for these purposes



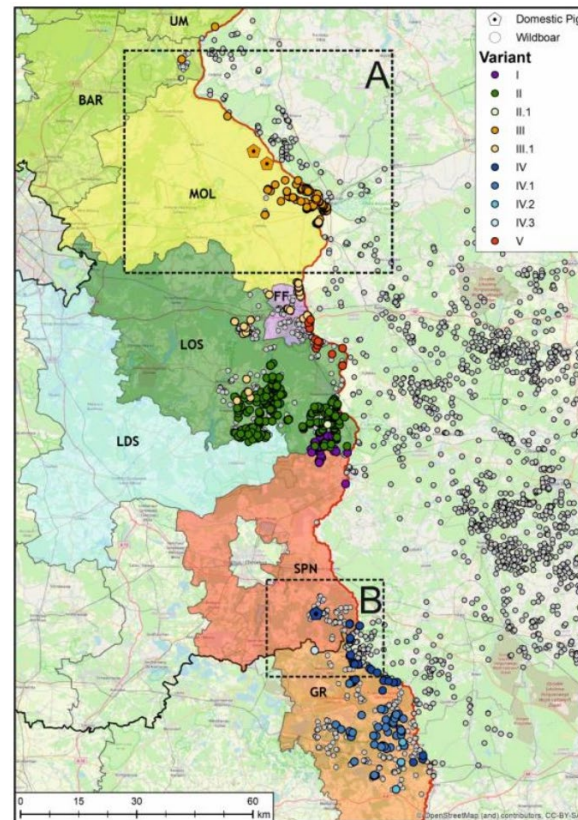
Alonso et al. *J. Gen. Virol.*, 99: 613–614 (mod.
From Dixon et al. *Virus Res.* 173 (1):3)



Whole genome molecular epidemiology: Germany



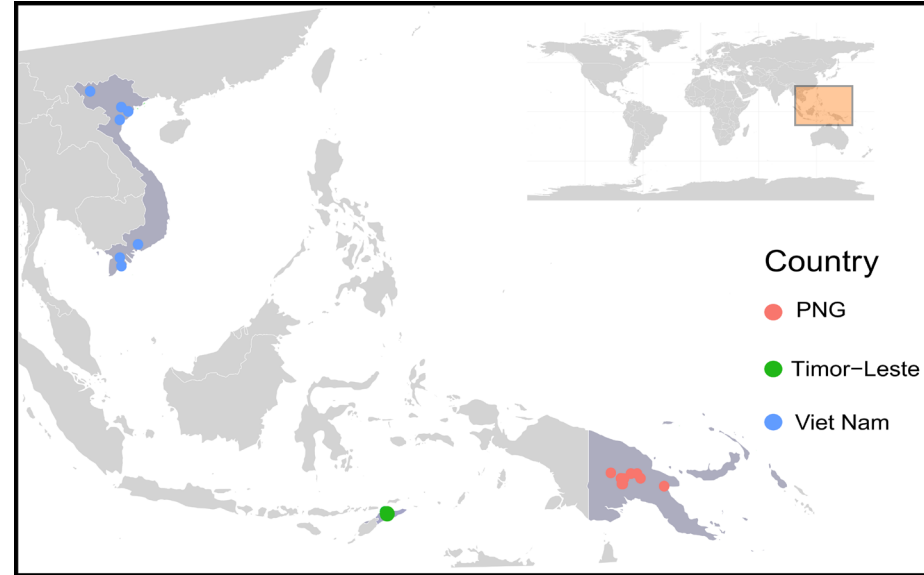
Forth et al. (2023) sequenced whole genomes and found small variations that were able to be used to *identify and track variants* along Germany's eastern border with Poland





ASFV Genomics: Emergence of microvariants of Genotype II in the Asia-Pacific region

- We investigated the presence of novel mutations in ASFV whole genomes from the Asia-Pacific
 - Derived from 25 samples collected from Viet Nam, Papua New Guinea, Timor-Leste
 - Submitted to ACDP ASF Reference Laboratory
 - Assessed usefulness of ASFV WGS for molecular epidemiology

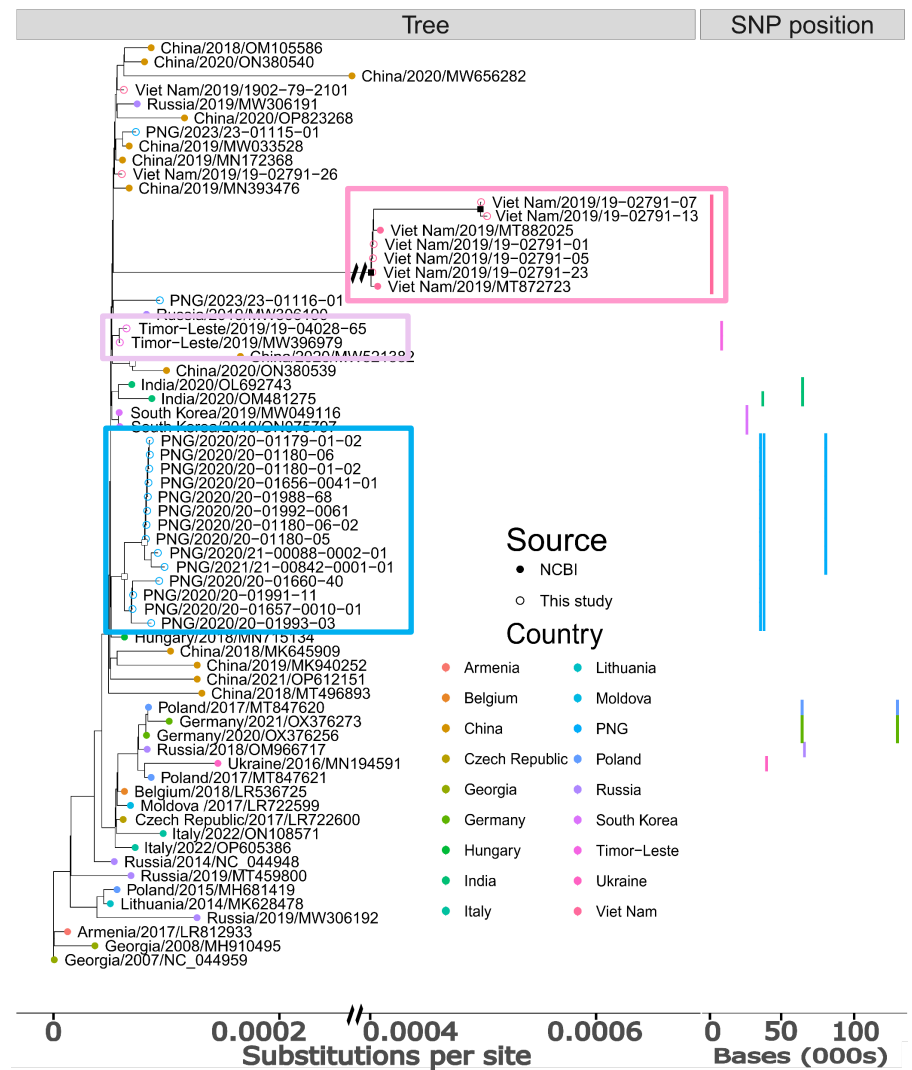


James O'Dwyer, Matthew Neave, ACDP



Emergence of microvariants in the Asia-Pacific

- Subtle genetic differentiation was found in samples from each locality
- Multiple individual mutations were found only within specific countries
- Geographic clustering of sequences from each country in phylogenetic analysis



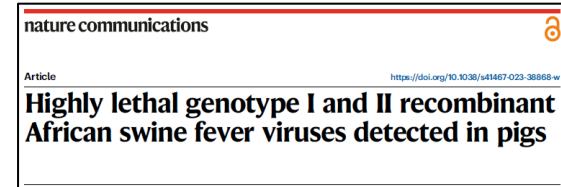


Detection of Recombinant ASFV

- Emergence of **recombinant genotype I/II** reported in China, Vietnam and E. Russia associated with acute ASF

- Identification by:

- Conventional PCR using 2 or more and sequencing *OR*
- WGS and bioinformatics → patterns of recombination across the genome



Dongming Zhao^{1,2}, Encheng Sun^{1,2}, Liayu Huang^{1,2}, Lilei Ding^{1,2}, Yuanmao Zhu^{1,2}, Jiwen Zhang¹, Dongdong Shen¹, Xianfeng Zhang¹, Zhenjiang Zhang¹, Tao Ren¹, Wan Wang¹, Fang Li¹, Xijun He¹ & Zhiguo Bu^{1,2}



Molecular Biology Reports (2024) 51:1011
<https://doi.org/10.1007/s11033-024-09961-0>





Which typing system is best to use?

- Single gene targets:
 - Original p72 partial gene typing offers simple and robust PCR-based method
 - Other targets provide additional levels of typing (e.g., *B602L* etc)
 - Lack sensitivity molecular epidemiology, phylogenetic analyses
- Whole genome sequencing:
 - Requires WGS and bioinformatics capability, and computing hardware
 - Sequencing from clinical samples is also technically challenging
 - WGS can provide highest resolution for molecular epidemiology and characterization of recombinant genomes



Summary

- Continued surveillance will be essential for understanding circulating types and genomic diversity of ASFV in the region
 - Additional whole genome sequences (as well as dissemination/publication) will enable analysis of evolution at the SEA regional level
- ACDP provides support for regional countries through the ASF Reference Laboratory, including for conventional typing, WGS and bioinformatics analyses
- New ASFV typing methods offer promise, but existing typing methods likely to be used alongside



Thank you

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