



# Comparison of the Sensitivity and Specificity of the Commercial Non-Structural Protein ELISA Kits Available in Thailand for Detecting the Antibodies to the Foot and Mouth Disease Virus

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## INTRODUCTION and OBJECTIVE

### INTRODUCTION

To control and prevent foot and mouth disease (FMD), the Non-Structural-Protein (NSP) ELISA test kit is used to detect antibodies to the NSP of FMD virus (FMDV) to distinguish between vaccinated animals and naturally infected animals with FMDV. Currently, various NSP ELISA test kits are available in Thailand. Therefore, the efficiency of the kits should be studied before selecting the appropriate kit.

### OBJECTIVE

This study aimed to compare the efficiency of detecting antibodies to NSP of FMDV using six commercial NSP ELISA test kits.

Table 1. Details of the characteristics of the NSP ELISA kits used in the study

| Test kit   | Manufacturer                              | Testing format    | Animals that can be tested                  | Positive interpretation criteria                                    |
|------------|---|-------------------|---|---|
| Biovet®    | Biovet Inc.,<br>Canada                    | Blocking<br>ELISA | cattle, buffaloes,<br>goats, sheep and pigs | The PI value depends on the status and type of animals.             |
| ID Screen® | IDvet,<br>France                          | Blocking<br>ELISA | cattle, buffaloes,<br>goats, sheep and pigs | S/N% ≤ 50%  |
| VDPro®     | MEDIAN Diagnostic<br>Inc.,<br>South Korea | Blocking<br>ELISA | cattle, buffaloes,<br>goats, sheep and pigs | S/N value ≤ 0.6   |
| IDEXX®     | IDEXX Laboratories<br>Inc.,<br>USA        | Blocking<br>ELISA | cattle, buffaloes,<br>goats, sheep and pigs | cattle, buffaloes, goats, sheep:<br>S/P% ≤ 35%,<br>pigs: S/P% ≤ 55% |
| PrioCHECK® | Prionics Lelystad<br>B.V.,<br>Netherlands | Blocking<br>ELISA | cattle, buffaloes,<br>goats, sheep and pigs | PI ≥ 50%  |
| KUcheck-F  | Kasetsart University,<br>Thailand         | Indirect<br>ELISA | cattle, buffaloes, goats<br>and pigs        | COD ≥ 0.2   |

**Note**  
 • %S/N = (OD test sample / OD negative control) × 100  
 • S/P ratio (sample to positive ratio) = (OD test sample - Average of OD negative control) / (Average of OD positive control - Average of OD negative control)  
 • Percent Inhibition; % PI = (OD<sub>450</sub> test sample) × 100 / OD<sub>450</sub> Max  
 • COD (corrected optical density) It is the measurement of light absorption.

- The 400 positive sera from cattle, buffalo and swine from the outbreak areas and the Bureau of Veterinary Biologics, the sera were confirmed for FMDV infection by ELISA Typing or RT-PCR.
- The 400 negative sera were brought from Japan and Australia, which have been certified FMD-free by WOA.
- The sera were used to examine the sensitivity and specificity of the test kits using ANOVA at 95% confidence level, concordance with Cohen's kappa analysis and diagnostic accuracy of the assay.



## MATERIAL and METHOD

## RESULTS and CONCLUSION

Table 2. The sensitivity and specificity values of the six NSP ELISA test kits.

| Test kit   | Animal type | Sensitivity (%) | 95% CI (SE) | Specificity (%) | 95% CI (SP) |
|------------|-------------|-----------------|-------------|-----------------|-------------|
| Biovet®    | Cattle      | 99.22 (383/386) | 97.75–99.84 | 99.00 (99/100)  | 94.55–99.97 |
|            | Pigs        | 100 (14/14)     | 76.84–100   | 100 (300/300)   | 98.78–100   |
|            | Total       | 99.25 (397/400) | 97.82–99.85 | 99.75 (399/400) | 98.62–99.99 |
| ID Screen® | Cattle      | 99.48 (384/386) | 98.14–99.94 | 100 (100/100)   | 96.38–100   |
|            | Pigs        | 100 (14/14)     | 76.84–100   | 100 (300/300)   | 98.78–100   |
|            | Total       | 99.50 (398/400) | 98.21–99.94 | 100 (400/400)   | 99.08–100   |
| VDPro®     | Cattle      | 97.67 (377/386) | 95.62–98.93 | 100 (100/100)   | 96.38–100   |
|            | Pigs        | 92.86 (13/14)   | 66.13–99.82 | 100 (300/300)   | 98.78–100   |
|            | Total       | 97.50 (390/400) | 94.45–98.79 | 100 (400/400)   | 99.08–100   |
| IDEXX®     | Cattle      | 97.93 (378/386) | 95.96–99.10 | 100 (100/100)   | 96.38–100   |
|            | Pigs        | 100 (14/14)     | 76.84–100   | 100 (300/300)   | 98.78–100   |
|            | Total       | 98.00 (392/400) | 96.10–99.13 | 100 (400/400)   | 99.08–100   |
| PrioCHECK® | Cattle      | 98.45 (380/386) | 96.65–99.43 | 100 (100/100)   | 96.38–100   |
|            | Pigs        | 100 (14/14)     | 76.84–100   | 100 (300/300)   | 98.78–100   |
|            | Total       | 98.50 (394/400) | 96.76–99.45 | 100 (400/400)   | 99.08–100   |
| KUcheck-F  | Cattle      | 98.96 (382/386) | 97.37–99.72 | 92.00 (92/100)  | 84.84–96.48 |
|            | Pigs        | 100 (14/14)     | 76.84–100   | 99.00 (297/300) | 97.11–99.79 |
|            | Total       | 99.00 (396/400) | 97.46–99.73 | 97.25 (389/400) | 95.13–98.62 |

Note: "Total" is calculated from the cumulative number of serum samples used in the testing.

Table 3. The Cohen's kappa statistics and diagnostic accuracy of the six NSP ELISA kits

| Test kit   | Cohen's kappa | 95% CI (Cohen's kappa) | Diagnostic accuracy (%) | 95% CI (Diagnostic accuracy) |
|------------|---------------|------------------------|-------------------------|------------------------------|
| Biovet®    | 0.99          | 0.98–1.00              | 99.50                   | 98.72–99.86                  |
| ID Screen® | 1.00          | 0.99–1.00              | 99.75                   | 99.10–99.97                  |
| VDPro®     | 0.98          | 0.96–0.99              | 98.75                   | 97.71–99.40                  |
| IDEXX®     | 0.98          | 0.97–0.99              | 99.00                   | 98.04–99.57                  |
| PrioCHECK  | 0.99          | 0.97–1.00              | 99.25                   | 98.37–99.72                  |
| KUcheck-F  | 0.96          | 0.94–0.98              | 98.12                   | 96.93–98.95                  |

### Discussion & Conclusion

Currently, there are commercially available NSP ELISA test kits in Thailand. The examination and evaluation of the effectiveness of these test kits is a key step in selecting the appropriate kit for the control and prevention of FMD in the country.

The study of six kits showed that the sensitivity was 97.50%-99.00%, specificity was 97.25%-100.00%, concordance was 0.96-0.99, and accuracy was 98.12%-99.75%. The results indicate that all six test kits were statistically similar and significant on sensitivity and specificity. In addition, the kits were found to have concordance and accuracy within the reliable criteria. This concludes that all kits could be used interchangeably. Therefore, the findings of this study can be used as decision-making data for selecting and purchasing a diverse range of effective kits.

### References

Brocchi, E., Bergmann, I.E., Dekker, A., Paton, D.J., Sammin, D.J., Greiner, M., et al. 2006. Comparative evaluation of six ELISAs for the detection of antibodies to the non-structural proteins of foot-and-mouth disease virus. *Vaccine*, 24: 6966–6979.

Department of Agriculture and Cooperatives, Ministry of Agriculture and Cooperatives. 2012. Agricultural Product Standards. MoAC. 10400-2012. Topic: Control Measures for Foot-and-Mouth Disease. 42 pages.

Doel, T.R. 2003. FMD vaccines. *Virus Res*. 91: 81–99. [http://dx.doi.org/10.1016/S0168-1702\(02\)00261-7](http://dx.doi.org/10.1016/S0168-1702(02)00261-7).

Geering, W.A. and J. Lubroth. 2002. Preparation of foot-and-mouth disease contingency plans. Emergency Prevention System, FAO, Rome.

Landis, J.R. and Koch, G.G. 1977. The Measurement of Observer Agreement for Categorical Data. *Biometrics*, 33: 159–174.

Linchongsabongkoch, W., Aunpomma, D. and Thongtha, T. 2004. The use of various non structural protein kits to differentiate between vaccinated and infected animals with foot and mouth disease virus. *J. Thai Vet Med. Assoc.* 55: 21–28.

Liu, W., Zhang, G., Yang, S., Li, J., Gao, Z., Ge, S., Yang, H., Shao, J. and Chang H. 2021. Development of a competitive chemiluminescence immunoassay using a monoclonal antibody recognizing 3B of foot-and-mouth disease virus for the rapid detection of antibodies induced by FMDV infection. *Virus Res*. 18: 193. doi: 10.1186/s12985-021-01663-4.

Mackay, D.K.J., Forsyth, M.A., Davies, P.R., Berlinzani, A., Belsham, G.J., Flint, M. and Ryan, M.D. 1998. Differentiating infection from vaccination in foot-and-mouth disease using a panel of recombinant, non-structural proteins in ELISA. *Vaccine*. 16 (5): 446–459.

World Organisation for Animal Health (OIE). 2019. "Principles and methods of validation of diagnostic assays for infectious diseases. Chapter 1.1.6." In Manual of Diagnostic Tests and Vaccines for Terrestrial Animals. OIE, Paris, France, 72–87. [Online]. Available: [www.oie.int/fileadmin/Home/eng/Health\\_standards/tahm/1.01.06\\_VALIDATION.pdf](http://www.oie.int/fileadmin/Home/eng/Health_standards/tahm/1.01.06_VALIDATION.pdf) Accessed November 19, 2022.

World Organisation for Animal Health (OIE). 2022. "CHAPTER 3.1.8. FOOT AND MOUTH DISEASE (Infection with foot and mouth disease)." [Online]. Available: <https://www.woah.org/en/disease/foot-and-mouth-disease/>. Accessed November 19, 2022.