

Singapore's Comments on the Report of the February 2020 Meeting of Aquatic Animal Health Standards Commission

Note: The parts 'double underline' and '~~strikethrough~~' were amendments drafted by the Code Commission, the parts highlighted **yellow** were amendments proposed at the Feb 2020 meeting and the parts in **green** are Singapore's proposed changes

Annex	Topic	Comments/Rationale for the amendment
11	<p>Infection with Spring Viraemia of Carp Virus (Chapter 2.3.9.) 3.5.2. <u>Preservation of Fixed samples for molecular detection</u></p> <p>Tissue samples for PCR testing should be preserved in 70-90% (v/v) analytical/reagent-grade (absolute) ethanol. The recommended ratio of ethanol to tissue is 10:1 based on studies in terrestrial animal and human health. The use of lower grade (laboratory or industrial grade) ethanol is not recommended. [Alternatives to ethanol can be mentioned if they can be referenced.]</p> <p>The material collected for virus culture is generally used for the molecular diagnostic assays, but additional tissue samples for RT-PCR can be preserved in commercially available RNA preservation solutions according to the manufacturers' recommendations, or, alternatively, samples can be preserved in 80-90%<u>90-95%</u> (v/v) analytical grade (absolute) ethanol at the recommended ratio of ethanol to tissue of 10:1.</p>	<p>We suggest to amend the percentage of ethanol from 80-90% to 90-95% (amendments highlighted in green) to be consistent as described in Section 4.5.1. Sample preparation and types, Chapter 2.3.0. General information OIE Manual of Diagnostic Tests for Aquatic Animals (14/11/2019 version):</p> <p><i>Alcohol-preserved samples:</i> in regions where the storage and shipment of frozen samples is problematic, 90–95% ethanol may be used to preserve, store, and transport certain types of samples. Pack for shipment according to the methods described above.</p>
11	<p>Infection with Spring Viraemia of Carp Virus (Chapter 2.3.9.) 3.6. Pooling of samples</p> <p>Traditionally pools of five animals have been used and more recently this has been increased to pools of ten animals for virus culture. However, no published data on the effect of pooling on test characteristics has been published.</p> <p><u>Pooling of samples from more than one individual animal for a given purpose</u></p>	<p>We suggest to include a recommended validation protocol for support of pooling.</p>

	<p><u>should only be recommended where supporting data on diagnostic sensitivity and diagnostic specificity are available. However, smaller life stages (e.g. fry) can be pooled to provide a minimum amount of material for testing.</u></p>	
<p>11</p>	<p>Infection with Spring Viraemia of Carp Virus (Chapter 2.3.9.)</p> <p>5. Test(s) recommended for surveillance to demonstrate freedom in apparently healthy populations</p> <p>The method for surveillance <u>of apparently healthy populations susceptible—fish populations</u> for declaration of freedom from infection with SVCV is inoculation of cell culture with tissue <u>homogenates extracts</u> (as described in Section <u>4.3.—4.5</u>) to demonstrate absence of the virus <u>or detection of SVCV RNA by conventional nested RT-PCR assays with tissue homogenate (as described in Section 4.4.2).</u></p>	<p>We suggest to include the amendments highlighted in green, to include conventional nested RT-PCR assay as one of the methods for surveillance of apparently healthy populations for declaration of freedom from infection with SVCV.</p> <p>According to “Table 4.1. OIE recommended diagnostic methods and their level of validation for surveillance of apparently healthy animals and investigation of clinically affected animals”, the rating of “Cell or artificial media culture” and “Conventional PCR” for “Surveillance of apparently healthy animals” is the same.</p>