Singapore's Comments on the Report of the September 2019 Meeting of Aquatic Animal Health Standards Commission

Note: The parts highlighted in green are Singapore's proposed changes.

Annex	Topic	Comments/Rationale for the amendment
10	**Infection with Spring Viraemia of Carp Virus (Chapter 2.3.9.)** **3.6. Pooling of samples** ~~Traditionally pools of five animals have been used and more recently this has been increased to pools of ten animals for virus culture. However, no published data on the effect of pooling on test characteristics has been published.~~ Pooling of samples from more than one individual animal for a given purpose should only be recommended where supporting data on diagnostic sensitivity and diagnostic specificity are available. However, smaller life stages (e.g. fry) can be pooled to provide a minimum amount of material for testing.	We would like to request for the SVC reference lab or experts to provide guidance or an assessment on the effect of pooling on diagnostic sensitivity and diagnostic specificity of the relevant tests, in support of the new position on pooling. The study on pooling of samples for AI surveillance, published by Spackman et al. (BMC Veterinary Research 2013, page 9:35) can be taken as an example.
10	**Infection with Spring Viraemia of Carp Virus (Chapter 2.3.9.)** **4.4. Nucleic acid amplification** **4.4.1. Real-time PCR** The following controls should be run with each assay: negative extraction control; positive control; no template control; internal PCR control if available and validated. Real-time RT-PCR assays are available to detect and confirm infection with SVCV (Yue *et al*., 2008; Zhang *et al*., 2009), however, they are not currently recommended as they have not been sufficiently validated. **4.4.2. Conventional PCR (PCR)** The following controls should be run with each assay: negative extraction control;	We suggest to include the amendments highlighted in green in 4.4.1 and 4.4.2 as internal PCR control may not always be available and validated.

	positive control; no template control; internal PCR control if available and validated. Nested reverse-transcription polymerase chain reaction (RT-PCR) (confirmation of virus identity from cell culture isolation or directly from fish tissue extracts)	